



## Detecting, bioaccumulating, and toxicological effects on aquatic trophic levels of nano plastic pollution

Manish Nandy<sup>1</sup>; Ahilya Dubey<sup>2</sup>; Dr. Monica Verma<sup>3</sup>

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### Abstract

The proliferation of micro- and nano-plastics (MNPs) in the ocean has become critical due to possible toxicological impacts on the environment, food web (FW) interactions, and human well-being. These plastic fragments originate from various sources, including degrading bigger plastic debris, goods, and emissions. This paper presents a comprehensive analysis of the distribution and hazards of MNPs in ocean habitats, their ecological conduct, and relationships within ocean food-webs, highlighting their harmful effects on ocean organisms. It examines the correlation between the size of particles and toxicity, their distribution throughout various organs, and the mechanism of transferring trophic status within the FW. Upon ingestion, MNPs are detected in multiple organs. The ingestion by the lowest-trophic-level creatures enables the advancement down the food chain (FC). This results in bioaccumulation and biomagnification, potentially damaging aquatic creatures' health, development, and actions. This study examines how MNPs, due to their persistence and metabolism, threaten marine ecosystems and disturb trophic interactions. The article discusses the ramifications of MNPs on human wellness, especially via ingesting contaminated seafoods, emphasizing the direct and indirect routes via which humans encounter these contaminants. The assessment underscores suggestions for further study, stressing the need for a combination of environmentally friendly and human wellness research to enhance risk evaluations and formulate mitigation plans to tackle the worldwide problem of plastic contamination in ocean ecosystems.

**Keywords:** Bioaccumulation, Toxicology, Nanoplastic pollution, Aquatic trophic levels

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1- Assistant Professor, Department of CS & IT, Kalinga University, Raipur, India.  
Email: ku.manishnandy@kalingauniversity.ac.in, ORCID: <https://orcid.org/0009-0003-7578-3505>

2- Research Scholar, Department of CS & IT, Kalinga University, Raipur, India.  
Email: ahilya.dubey@kalingauniversity.ac.in, ORCID: <https://orcid.org/0009-0008-1681-8823>

3- Professor, New Delhi Institute of Management, New Delhi, India. Email: monica.verma@ndimdelhi.org,  
ORCID: <https://orcid.org/0000-0003-2789-1117>

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## Introduction

Plastics are now essential to our lives; however, their incorrect disposal and subsequent fragmentation or destruction have led to a rise in the environment's micro- and nanoplastics (MNPs) (Zhu *et al.*, 2023). This includes both primary MNPs, which are deliberately manufactured at micro- and nanoscale levels, and added MNPs, which arise from the decomposition of larger plastic products (Vinusha *et al.*, 2024). The appearance of these minuscule plastic particles has elicited worries over their ability to accumulate in aquatic environments and potentially have detrimental impacts on marine species (Shah and Bansalm, 2023). Their contamination has emerged as a significant worldwide issue, with its adverse effects on the environment and individuals being broadly recognized. The impact of Microplastics (MPs) (Jin *et al.*, 2022) is well-established, although the existence and possible dangers of MNPs in the natural world have recently attracted considerable scrutiny. MPs, defined as particles of plastic smaller than five millimeters, and Nanoplastics (NP) (Torres-Ruiz *et al.*, 2021), weighing less than 100 nm, have emerged as ubiquitous pollutants in aquatic environments, posing potential threats to marine creatures and the general health of these environments.

The marine environment is an essential habitat for various creatures, and any disturbance caused by MNPs might have significant repercussions (Dube and Okuthe, 2023). A thorough examination of the toxicological consequences of MNPs, including

exposure assessment, possible toxicity, and subsequent transmission through the food web (FW), is essential. Grasping these elements is crucial for recognizing the comprehensive impacts of MNPs on aquatic environments (Pokric *et al.*, 2015). This entails investigating the relationships physically, chemically, and biologically among plastics and ocean life, emphasizing how these particulates influence diverse marine creatures' development, reproduction, conduct, immune response, and general wellness, from miniature plankton to larger entities like fish and whales (Agarwal and Yadhav, 2023; Rajak *et al.*, 2024).

Examining the transportation of MNPs via the food chain (FC) is essential for comprehending the wider environmental consequences (Escobedo *et al.*, 2024). Primary producers, like phytoplanktons, can absorb and gather MNPs. Such MNPs are ingested by tiny creatures such as zooplanktons and transmitted to the highest trophic areas (Provencher *et al.*, 2022). The transport of MNPs throughout the alimentary web raises issues over potential growth and biomagnification of these fragments, resulting in heightened exposure for apex organisms, especially humans.

The hazards posed by MNPs within FWs are a focal point in the paper concerning the toxicological impacts of MNPs. Various researchers have sought to comprehend how the existence of these polymers can disturb the FW's equilibrium and operation. This alteration induces cascading consequences on the aquatic environment factors, encompassing changed population patterns, modifications in diversity, and possible harm to biodiversity. The vital

transmission of MNPs to people via seafood intake raises concerns regarding health implications, necessitating a thorough study (Vishaka and Selvi, 2017).

## Background

### *Preparations and Purifications of Palladium-imposed NP*

Nanospheres encapsulating palladium were synthesized using a previously documented process with some modifications. Sodium Dodecyl Sulphates (SDS, 0.2 g) and acrylonitriles (ACN, 7.5 mL) are generally introduced into a cylindrical container holding 45 mL of water, under a nitrogen environment. This mixture included 5 mL of a 0.12 mol L<sup>-1</sup> potassium persulfate (KPS) mixture with rapid agitation. Over 5 minutes, 6 mL of 4-nonylphenyl 3-sulfopropyl ether and 10 mL of K<sub>2</sub>PdCl (0.06 mol L<sup>-1</sup>) liquids were introduced into the vessel using a syringe pump at a consistent rate, followed by a maintenance period at 65 °C for 40 minutes. Afterwards, an additional 0.25 g of KPS was delivered straight into the apparatus in 5 mL of pure water, followed by the injection of a mixture comprising 25 mL of H<sub>2</sub>O, 10.5 mL of plastic, and 0.5 g of divinylbenzene into the reaction vessel at an injection rate of 0.06 mL<sup>-1</sup>. The solution was sterilized with nitrogen at ambient temperature for 24 hours to eliminate residual styrene and acrylic acid, removing over 99% of the volatile chemicals. The dimensions and morphology of the produced Pd-imposed NP were analyzed using Transmission Electron Microscopy (TEM), Scanning Electron Microscopy (SEM), and Dynamic Light Scattering (DLS) (Tarrés

*et al.*, 2022). The long-term stability of Pd-imposed NP was assessed by measuring the dissolution of palladium through ultrafiltration.

### *Fabrication of Aqueous Microcosm*

The protocol established an aquatic microcosm comprising water, sand, and biota in a glass tank of 40 cm long, 45 cm in breadth, and 30 cm in height. The biota species and their respective quantities were established based on prior research concerning freshwater environments, with some adjustments. All animal studies complied with the handling and use standards for laboratory animals and received approval from the Animal Ethics Board. These microbes are prevalent species in China's lakes and rivers, forming a natural FC in Eurasia. The tiny organism was permitted for two months with a brightness of 3000lux and a 16/8 hour sunlight at 25/16 °C before NP treatment. The charcoal-filtering water supply was refilled biweekly to compensate for losses from evaporation.

### *Exposure of the Manufactured Miniature to Pd-Doped NP*

The miniature NP therapy was administered to both pulse and continuous patients. Pulse-treatment miniatures were administered a single dosage of 15 mL NP solution, equating to 196 mg of NPs introduced into a mean of 24 L of microscopic H<sub>2</sub>O. This resulted in an ending aqueous level of 8.7 mg/L. No specific levels or anticipated levels of NP in aquatic environments have been documented (Bathi *et al.*, 2021). An 8.7 mg L<sup>-1</sup> was selected based on research, which indicated a cumulative level varying from 0.2 to 120 mg L<sup>-1</sup> of NP.

Chronic-treatment microcosms were administered a weekly dosage of 3 mL NP suspensions (15 mg mL<sup>-1</sup>) for 2 months, resulting in an accumulated NP dosage equivalent to that of the pulsing therapy category. The controlling replicas were supplied exclusively with charcoal-filtered municipal water. Every therapy comprised three replicate miniatures. Following a 49-day contact, every specimen was collected, measured, and processed, while the particles were sun-dried and pulverized using a spatula.

#### *Processing and NP Characterization*

Each specimen received acid digestion before NP measurement. In summary, sediments (about 0.3 g), biotic (0.2-0.6 g), and H<sub>2</sub>O (1 mL) specimens were combined in a 12 mL poly-propylene capacity with 3 mL of water from the region, and the resultant combination was warmed at 95°C for 24 hours. After the digestive process, the mixture was vaporized to approximately 0.6 mL, then diluted with purified H<sub>2</sub>O to a final volume of 15 mL. The Pd content was measured using an Agilent 7800 Inductively-Coupled Plasma-Mass Spectrometry (ICP-MS). The ionic platinum standard mixture (1100 mg L<sup>-1</sup>) and rhodiums (Rh) reference solutions (12 mg L<sup>-1</sup>) were diluted to serve as the measurement mixture and inner condition, correspondingly.

#### *Quality Assessment (QA) /Quality Controlling (QC)*

All features were meticulously cleaned with pure H<sub>2</sub>O to reduce Pd backgrounding stages, and specific measures were taken with contact, collection, and analysis. Every canister was constructed from polyamides. The

recuperation rates of NP in various sample environments were examined by stimulating control specimens with calibrating fluids. Process samples were examined to detect contamination. Simultaneously, procedure duplicates and spiked specimens were evaluated within each collection of about fifteen specimens during therapy, and a calibrating condition was conducted periodically (approximately every 15 samples) to assess instrumental drift. The threshold of identification and Limits Of Quantitation (LOQs) were established by evaluating the quantities corresponding to Signals-to-Noise Ratios (SNR) of 4 and 9 in blank samples. A calibrating curve was established across diverse sample matrices to guarantee the accuracy of NP tenacity. NP's average recovery rate (n = 4) from spiked H<sub>2</sub>O, sand, and biotic varied from 75.2% to 98%, 71.3% to 83.7%, and 76.4% to 99.6%, with comparative variances < 9.2%. The limits of determination and LOQs are 0.04mgL<sup>-1</sup> and 0.2mgL<sup>-1</sup> in H<sub>2</sub>O, 0.5mgkg<sup>-1</sup> and 1.5mgkg<sup>-1</sup> in debris, and 0.5-0.8mgkg<sup>-1</sup> and 1.5-2.8mgkg<sup>-1</sup> in organisms.

#### **Detecting, Bioaccumulating, and Toxicological Effects**

##### *Characterization of NP*

Pd-doped NP exhibiting a 'raspberry' -like morphology and a median dimension of 105 ± 12 nm were utilized in this investigation. The power dispersive X-ray spectrometer mapping observations in palladium were primarily localized in the center of the NP, corroborating the findings. DLS research indicated that the Pd-imposed NP exhibited an adverse charged surface (zeta voltage -37.2 ±

1.2mV) with a hydro-dynamic radius of  $150 \pm 2$  nm. To ascertain the Pd in the MNPs, serial solutions of NP from 0.1 to  $1100\text{mgL}^{-1}$  were examined using ICP-MS. The Pd content exhibits a linear relationship with that of NPs, constituting 0.9% of the overall mass of MNPs.

Pd leaching was quantified under diverse settings for a specified duration to assess Pd's NP stability. Merely 0.5% of the ionic palladium from the total palladium was not integrated into the NP during synthesis. No elevation in the Pd was detected in the  $\text{H}_2\text{O}$  from the faucet when exposed to light. Since stomach juice and stool fluid are two critical biological liquids that could substantially influence metal durability due to their pH values being beyond the range of neutral, the long-term stability of Pd was evaluated in these two modeled waters. The findings indicated that the soluble palladium did not surpass 3.6% in the synthetic liquid and fluid after being agitated for 49 days. The results showed the exceptional stability of Pd in nanoparticles, implying that Pd serves as a surrogate for NP. Previous research has overlooked the leaching of biological material from NPs due to the remarkable stability of polymers under benign circumstances, despite the potential release of minimal quantities in exposition.

#### *Mass Balancing*

The manufactured Pd-imposed NP were introduced into the miniatures using one or more dosage treatments, achieving equivalent cumulative levels. Water quality substantially influences NP's association and migratory characteristics; standard water quality variables such as pH, temperature, and Dissolved Oxygen

(DO) in the replicated miniatures were assessed. The water characteristics of the replicated miniatures infused with NP were consistent with those of the untreated group over the 49-day contact, indicating that NP exposures had minimal effect on the circumstances of the artificial ecosystem. The amounts of NP in the water column and particles were measured. NP floating in water sank swiftly, resulting in higher levels in the sand following an initial spike for both continuous and pulse groups receiving therapy. After two days, the NP attained half equilibrium in  $\text{H}_2\text{O}$  ( $4.3\text{mgL}^{-1}$ ) and debris (approximately  $25\text{mgkg}^{-1}$ ) within the sample. Despite the prevalence of plastic trash floating in the  $\text{H}_2\text{O}$  column due to its small weight, the findings revealed a substantial accumulation of NP in the sediment.

Their physical and chemical characteristics change significantly when big material breaks into NP, leading to modifications in their settlement behavior, particularly due to heteroaggregation with suspended debris in natural waterways. The settling characteristics of NPs resembled those of metallic NPs in the surroundings, suggesting that NPs provide a greater risk in sedimentation compared to aquatic environments.

The NP distribution in the replicated miniatures was assessed following a 49-day exposure period. The recovery rates of the total spiking NP ranged from 87.1% to 94.5% in the built environment, demonstrating negligible loss of NP during exposure. NP were allocated between every space in the modeled miniatures, with the quantity retained exhibiting significant variability among

these spaces. Most overall recoverable NPs were found in the debris, including  $93.1 \pm 8.5\%$  for the chronic-treatment sample and  $93.1 \pm 4.5\%$  for the treating category. This indicates that soil is the primary storage of MNPs in the natural world. The proportion of MNPs in the organisms was significantly minimum compared to the sand. Approximately 3.4% and 0.5% of the overall NP were detected in the biotic for the persistent and pulse treatments, indicating a greater bioaccumulating ratio of NP during prolonged exposure. Approximately 2.0–3.5% of NP floating in the liquid stage were seen after 49 days of exposures, indicating their potential for traveling long distances in aquatic environments.

#### *Bioaccumulation and Diffusion of NP in Microbes*

The NP levels varied from 4.7 to 178  $\mu\text{g g}^{-1}$  and from 3.4 to 76.2  $\mu\text{g g}^{-1}$  wet mass across all biota specimens. The bioaccumulation of NP in animals is influenced by multiple factors, such as depuration capacity and feeding conduct, rather than solely by their surroundings. The mean Bioaccumulation Factor (BAF) in organisms, determined as the rate of the observed amount of NPs in ocean animals to that in fluid, was between 28.1 and 1080 for the chronic-treatment sample and 11.4 and 290 for the pulse-treatment category, correspondingly. The findings were below the standard for BAF substances, indicating that NPs are not BAF. The evaluation of average NP levels demonstrated that human burden principles in the grouping were considerably bigger, suggesting that NP metabolism was contingent upon ongoing or pulse-exposure conditions.

The most significant amounts in crops for continuous and pulse regimens were observed, attributable to their bioaccumulation functions, environments, physiological research, and feeding behaviors.

#### **Identified Research Deficiencies and Prospective Directions**

This research primarily seeks to synthesize existing knowledge regarding MNPs in aquatic environments. It examines their primary sources, the routes by which they infiltrate the FW, and their toxic impacts on various ocean creatures, affecting human wellness. The results highlight multiple opportunities for studies that might be pursued to enhance comprehension of the prevalence and implications of MNPs in marine environments.

1. Numerous studies examining the impacts of MNPs on aquatic creatures, notably in vitro and in vivo studies, employed extremely brief exposure periods. However, the duration required for MNPs to impact these organisms remains unclear. This does not entirely capture the intricacies of aquatic ecological situations, where numerous elements related to these polymers can exert significant effects. Subsequent research evaluating the impacts of MNPs, emphasizing prolonged exposure durations, should thoroughly investigate the chronic and accumulated consequences. Comprehending the persistence and accumulation of these particles throughout time and their interactions with other creatures in the environment would provide significant viewpoints for future research.

2. Although prior studies indicate that MNPs are conveyed to cells via extracellular fusions and blood flow, a thorough examination of the transferring situations and pathways across different organs remains inadequate. The absorption of MNPs relates to biological factors and processes that occur in vivo. The current literature primarily concentrates on laboratory transmission studies without incorporating diverse aquatic creatures (e.g., some studies exclusively utilized fish, which do not accurately reflect transfer handles in other varieties). In contrast, reptiles, mammals, and amphibians display significant digestive, breathing, and bloodstream differences. Expanding the laboratory modeling animal framework is crucial for examining the translocation mechanisms of MNPs in different species.

3. Recent investigations indicate that chemical and biological processes can efficiently eliminate MNPs in aquatic environments. MNPs have not been thoroughly investigated regarding their deterioration. To deliver efficient and pragmatic solutions for the global pollution problem caused by MNPs, additional thorough studies on MNP degrading methods will be conducted in the future.

4. A comprehensive study is necessary to identify elements that could substitute for plastics to mitigate pollution and the toxicological impacts of NP completely.

5. MPs function as carriers for diverse environmental pollutants. Although research has underscored the possible hazards associated with these pollutants, examining the harmful effects of compound contaminants across each

trophic level remains essential. Subsequent research should focus on pollutants' connection, adhesion, and metabolism without MNPs.

6. The harmful effects of MNPs have been examined in-vitro and in-vivo; knowledge regarding higher-level microbes remains limited, and investigating MNPs at the cell and physiological level poses challenges. Future studies explore these concerns.

## Conclusion

This article offers a comprehensive analysis of the detrimental effects of MNPs on aquatic animals, including impaired development, tissue damage, oxidative stress induction, DNA damage, and reproductive harm. The reduction in the size of MPs over time enhances the area of their surfaces, potentially allowing for greater absorption of pollutants, which poses considerable health concerns to animals and human wellness via the FC. The ocean surroundings, essential for the sustenance and refuge of fish prey, enhance their total population. The surrounding area's pollution jeopardizes food safety and threatens human health via the FC. Although research on MNPs has progressed considerably in recent years, there is a necessity for more sophisticated and comprehensive toxicology methodologies to fully understand the role of MNPs and address the adverse effects resulting from their accumulation in the FC or FW. Ongoing discourse and ambiguity exist over the consequences and implications of MNPs upon ingestion by ocean organisms and people. The limited study on the trophic dissemination of MNPs pollution hinders the synthesis of results, especially with

risks to human health via effective movement within the FW. Considerable gaps remain, underscoring the need for additional study to elucidate the consequences of MNPs in the ocean environment and delineate future study trajectories. Formulating effective reduction strategies is essential for protecting the environment and human welfare against the rising hazards presented by MNPs.

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